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# Annular pancreas in Trichorhinophalangeal syndrome type II with 8q23.3-q24.12 interstitial deletion

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#### **Abstract**

**Background:** Trichorhinophalangeal syndrome type II (TRPS II, OMIM # 150230) is a rare autosomal dominant genetic disorder characterized by craniofacial and skeletal abnormalities. Loss of functional copies of the *TRPS1* gene at 8q23.3 and the *EXT1* gene at 8q24.11 are considered to be responsible for the syndrome.

**Case Presentation:** Herewith, we report an 8-year-old girl with sparse scalp hair, bulbous nose, thin upper lip, broad eyebrows, phalangeal abnormalities of both hands/toes, multiple exostoses, mild intellectual impairment and severe malnutrition. In addition, the patient also had annular pancreas, a rare co-existing feature in patients with TRPS II.

**Conclusions:** A contiguous 5.47 Mb deletion involving 8q23.3-q24.12 was detected by array comparative genomic hybridization (aCGH), leading to haploinsufficiency of 10 protein coding genes, 1 long non-coding RNA and 1 microRNA. Quantitative PCR (qPCR) examination confirmed half-reduced DNA copy of the patient and normal expression of both parents, indicating a *de novo* origin of the deletion and complete penetrance of the mutation.

Keywords: Annular pancreas, 8q23.3-q24.12 deletion, Trichorhinophalangeal syndrome type II, Chinese, qPCR

#### **Background**

Trichorhinophalangeal syndrome type II (TRPS II), also known as Langer–Giedion syndrome (LGS), is a contiguous gene deletion syndrome involving loss of functional copies of multiple genes on 8q24.1. Hall et al. first described this disorder in 1974 [1], which combines the clinical features of trichorhinophalangeal syndrome type I (OMIM # 190350) and multiple exostoses type I (OMIM # 133700). Affected individuals are characterized by multiple dysmorphic facial features including large, laterally protruding ears, a bulbous nose, elongated upper lip, sparse scalp hair, winged scapulae, multiple cartilaginous exostoses, and mental retardation. Although familial cases have been occasionally observed [2], most reported cases to date are sporadic ones. In 1984, Buhler and colleagues identified an interstitial deletion involving chromosome

8q24 in an individual with TRPS II [3], and later on proposed this complex disease as a chromosome deletion syndrome involving both TRPS1 and the gene for multiple hereditary exostoses EXT1. Patients with TRPS II often show clinical variability depending on the loss of additional genes in the deleted region [4]. For example, mild to moderate intellectual disability, conductive hearing loss, epilepsy, congenital nephrotic syndrome, growth hormone deficiency, and a submucosal cleft palate have all been described as additional features of TRPS2/LGS depending on the deletion size [5–9].

Here, we present an 8-year-old female with sparse scalp hair, facial dysmorphism, malaligned crowded teeth, multiple exostoses, winged scapulae, annular pancreas, mild intellectual impairment and an 8q23.3-q24.12 deletion detected by aCGH.

#### Case presentation

Our female patient was the first child of healthy unrelated parents of normal intelligence. She was born by caesarian section at 40 weeks of gestation after an

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Fig. 1 Clinical features of our patient diagnosed with trichorhinophalangeal syndrome type II: a, long and clubbed-fingers. a, b Nails of fingers and toes are typically thin and dystrophic

uneventful pregnancy. Her birth weight was 3050 g ( $<50^{th}$  centile), length 49 cm ( $<50^{th}$  centile) and head circumference 33 cm ( $<50^{th}$  centile). Both parents were normal, and there was no family history of congenital malformations. The patient was referred to our Department of Pediatric Surgery at 8 years of age with recurrent vomiting and severe malnutrition. On clinical

examination, the girl had brachycephaly, fine and sparse hair, a bulbous tip of the nose, long and flat philtrum, thin upper lip, broad eyebrows, malaligned crowded teeth, micrognathia, microcephaly (with a head circumference of 42 cm at the age of 8-year-and-5-month), hypotonia and hyper-extensibility of the joints. In addition, she had winged scapulae, long and clubbed-

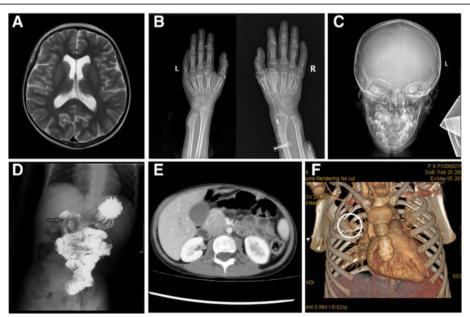
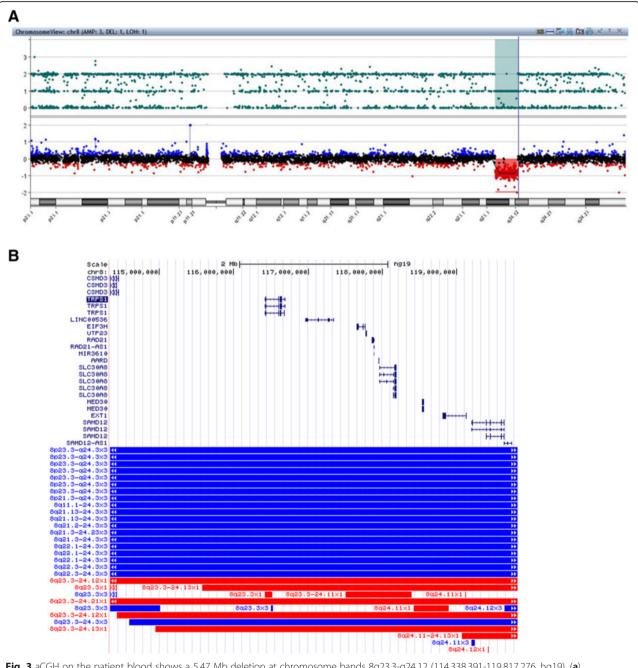


Fig. 2 Imaging examinations of the patient. a Brain MRI showed encephalatrophy-like changes. b, c X-ray examination showed short metacarpals of the fifth finger, cone-shaped epiphyses of the proximal inter-phalangeal joint of middle phalanx of the fingers (white arrow), multiple exostoses (white arrow head) and malaligned crowded teeth. d Upper GI series showed insufficient distention at the pylorus and duodenum (black arrow) indicating encircled pancreatic tissue. e Axial CT image clearly showed annular pancreatic tissue encircling the second portion of the duodenum (white circle). f CT volume rendering revealed exostoses (white arrow head) and fusion malformation of the second and third ribs on the right side (white circle)

fingers. Her finger and toe nails were typically thin and dystrophic (Fig. 1). Her weight was only 13.25 kg (<1st centile), and her height was 120 cm (<15th centile). She showed mild intellectual impairment and psychomotor retardation. Heart and abdominal ultrasound revealed no pathological abnormalities. Brain MRI showed encephalatrophy-like changes (Fig. 2, a). X-ray examination showed short metacarpals of the fifth finger, coneshaped epiphyses of the proximal inter-phalangeal joint

of middle phalanx of the fingers and malaligned crowded teeth (Fig. 2, b and c). Upper GI (gastrointestinal) series showed insufficient distention at the pylorus and duodenum indicating encircled pancreatic tissue. Axial CT image clearly showed annular pancreatic tissue encircling the second portion of the duodenum (Fig. 2, d and e). CT volume revealed exostoses of the scapulae and fusion malformation of the second and third ribs on the right side (Fig. 2, f). Informed



**Fig. 3** aCGH on the patient blood shows a 5.47 Mb deletion at chromosome bands 8q23.3-q24.12 (114,338,391-119,817,276, hg19). (a) Chromosomal view and (b) UCSC zoom in view

consent and permission of publication of identifiable photographs of the patient have been obtained.

#### Results

Cytogenetic analysis at a resolution of 400 bands displayed a normal female karyotype (46, XX). Wholegenome aCGH analysis on the patient detected a 5.47 Mb deletion at chromosome bands 8q23.3–q24.12 (114,338,391–119,817,276, hg19). The deleted region encompasses altogether 10 protein coding genes (CSMD3, TRPS1, EIF3H, UTP23, RAD21, AARD, SLC30A8, MED 30, EXT1 and SAMD12), 1 long non-coding RNA (LINC00536) and 1 microRNA (MIR3610) (Fig. 3). Targeted gene sequencing revealed no pathological mutation of TRPS1 and EXT1 gene in the patient (data now shown). Quantitative PCR tests show approximately half-reduced DNA copy of the patient and normal expression of both parents, indicating a de novo origin of the deletion (Fig. 4).

#### Discussion

Here we report the identification of a novel deletion of 8q23.3–q24.12 responsible for a Chinese patient suffering from Trichorhinophalangeal syndrome type II. Quantitative PCR confirmed the heterozygous variant which leads to haploinsufficiency of 10 protein coding genes, 1 long non-coding RNA and 1 microRNA. Both parents show a normal expression level as control, indicating a *de novo* origin of the deletion and complete penetrance of the mutation.

The trichorhinophalangeal syndrome (TRPS) is a rare autosomal dominant disorder characterized by cranio-facial dysmorphism and bone deformities. Depending on the severity of the phenotype, three subtypes have been described: TRPS I (OMIM # 190350) and TRPS III (OMIM # 190351) are caused by mutations in TRPS1; and TRPS II (OMIM # 150230) is caused by a contiguous gene deletion affecting TRPS1 and EXT1 amongst others. Diagnosis of TRPS is normally based on clinical features and radiographic examination [10]; however, genetic analysis is often helpful especially in

the case of non-classical clinical presentation [11, 12]. To date, more than 100 patients with TRPS have been reported in the literature [5], and features common to all three subtypes include sparse, slowly growing scalp hair, laterally sparse eyebrows, pear-shaped nose and large, protruding ears. Further typical characteristics include a long flat philtrum and a thin upper lip. In addition, short hands (feet) and impaired growth are common in all types of TRPS, while body weight is usually normal in relation to height. With respect to the head circumference, it is typically normal throughout life, except for those with a genomic deletion in whom one-third could have head circumference below the 3<sup>rd</sup> percentile [13].

The most distinguishable feature for TRPS II from type I and III is exostoses, which are only present in those individuals with a deletion that extends to EXT1. Furthermore, intellectual disability (developmental delay) is present in most but not all TRPS II patients, typically mild to moderate in severity, and does not seem to correlate with the size of the deleted segment. Maas et al. summarized the information on 103 cytogenetically or molecularly confirmed TRPS individuals, and suggested that the size of contiguous gene deletions vary considerably in TRPS II and there are no common breakpoints [5, 11]. In our patient, aCGH analysis revealed the deletion of both TRPS1 and RAD21 genes which is in accordance with the dysmorphic facial features. Multiple exostoses, winged scapulae, impaired growth and mild intellectual impairment highly support the clinical diagnosis of TRPS II. Interestingly, the patient was also found to have annular pancreas, which, to our knowledge, is a novel co-existing feature in patients with TRPS II. Annular pancreas is a relatively rare congenital anomaly [14] characterized by an extension of pancreatic tissue around the second part of the duodenum and is thought to represent aberration in the development of the ventral pancreatic bud [15]. Recent studies highlight the role of the hedgehog signaling pathway in the development of this anomaly [16]. Overexpression of the ventral-specific gene transmembrane 4

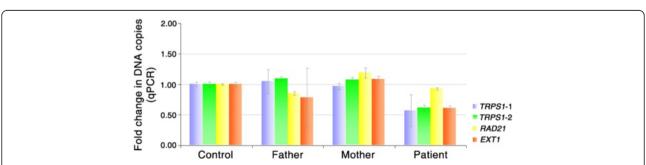


Fig. 4 qPCR examinations show approximately half-reduced DNA copy of the patient and normal expression of both parents, indicating a *de novo* origin of the deletion

superfamily member 3 (*tm4sf3*) has also been associated with annular formation [17]. Moreover, isolated case reports of familial annular pancreas have also been documented, suggesting a genetic basis for the development of this anomaly [18]. The 8q deletion in our patient spans 5.47 Mb and none of the genes within this region seemed to be related to the hedgehog signaling pathway. Further study is warranted to provide an answer to the underlying pathological mechanism.

#### Conclusions

In summary, we present molecular cytogenetic characterization of a *de novo* deletion involving 8q 23.3–q24.12 in an 8-year-old female with clinical features of TRPS II and annular pancreas. This is the first molecularly diagnosed patient with typical TRPS II in China. Our findings further confirm the relationship between TRPS II and 8q24.1, and expand the phenotypic and molecular spectrum of the disorder.

#### Materials and methods

#### array comparative genomic hybridization (aCGH)

DNA from peripheral blood was isolated by the Blood and Tissue kit (Qiagen, Valencia, CA). Whole-genome aCGH on patient's blood was performed using Agilent CNV + SNP 180 K oligonucleotide array (Agilent Technologies Inc., Palo Alto, CA). This array has 170,334 probes and the overall median probe resolution is 11–13 kb.

#### Quantitative polymerase chain reaction (qPCR)

Quantitative polymerase chain reaction (qPCR) was carried out in the presence of SYBR Green measuring the fluorescence signal produced by the binding of SYBR Green to the studied amplicons and compared with a reference sample. Average copy numbers of GAPDH in normal candidates (copy numbers = 2) were used as endogenous control. Real-time qPCR was performed with 7500 Fast Real-Time PCR System (Life, USA). The copy numbers of TRPS1, RAD21 and EXT1 were calculated by using the comparative C (T) method. The qPCR program was as follows: start at +50 °C for 2 min, then at +95 °C for 10 min, followed by 40 twostep cycles (15 s at +95 °C, 1 min at +60 °C). Reactions were adjusted to a final volume of 20 ul/well using 2 ul of genomic DNA (10 ng/ul), 1 ul of both of primers (10 pmol/ul), 6 ul of sterile water and 10 ul of Master Mix. The primers for GAPDH are (forward) 5'-AATG GGCAGCCGTTAGGAAA-3' and (reverse) 5'-AAAAG CATCACCCGGAGGAG-3'. The primers for target genes are as following: TRPS1-1 (forward) 5'-AAG-CACTCTAAAGCCACATGC-3' and (reverse) 5'-TGT GCCAAGAACTTTTCCAG-3'; TRPS1-2 (forward) 5'-AATGCAAATGGCGGATATGT-3' and (reverse) 5'- CCAGGCCAACACTGCTTTAT-3'; *RAD21* (forward) 5'-TGGGCCTGATAGTCCTGATT-3' and (reverse) 5'-GCTCCAATGCAAATGCTTCT-3'; *EXT1* (forward) 5'-TGTGACAAAATTGCCTCCAA-3' and (reverse) 5'-GCTGAACCACCAGTGAGTGA-3'. Statistical analysis was performed using Fisher's exact test. *P* < 0.05 was considered statistically significant.

#### Consent

Written consent was obtained from the patient and her parents for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

#### Abbreviations

aCGH: Array comparative genomic hybridization; LGS: Langer–Giedion syndrome; qPCR: Quantitative polymerase chain reaction; TRPS II: Trichorhinophalangeal syndrome type II.

#### Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

LS and KY extracted DNA. QL, XC and QJ performed array-CGH, and draft the manuscript. HX carried out quantitative PCR. QL, ZZ, YY, PX and ZG contributed to clinical evaluation of the family. WC helped writing and editing the manuscript. QJ and LL supervised the whole work. All authors read and approved the final manuscript.

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#### References

- Hall BD, Langer LO, Giedion A, Smith DW, Cohen Jr MM, Beals RK, et al. Langer-Giedion syndrome. Birth Defects Orig Artic Ser. 1974;10(12):147–64.
- Ludecke HJ, Wagner MJ, Nardmann J, La Pillo B, Parrish JE, Willems PJ, et al. Molecular dissection of a contiguous gene syndrome: localization of the genes involved in the Langer-Giedion syndrome. Hum Mol Genet. 1995;4(1):31–6.

- Buhler EM, Malik NJ. The tricho-rhino-phalangeal syndrome(s): chromosome 8 long arm deletion: is there a shortest region of overlap between reported cases? TRP I and TRP II syndromes: are they separate entities? Am J Med Genet. 1984:19(1):113–9.
- Ludecke HJ, Schmidt O, Nardmann J, von Holtum D, Meinecke P, Muenke M, et al. Genes and chromosomal breakpoints in the Langer-Giedion syndrome region on human chromosome 8. Hum Genet. 1999;105(6):619–28.
- Maas SM, Shaw AC, Bikker H, Ludecke HJ, van der Tuin K, Badura-Stronka M, et al. Phenotype and genotype in 103 patients with tricho-rhino-phalangeal syndrome. Eur J Med Genet. 2015;58(5):279–92.
- Lu FL, Hou JW, Tsai WS, Teng RJ, Yau KI, Wang TR. Tricho-rhino-phalangeal syndrome type II associated with epiglottic aplasia and congenital nephrotic syndrome. J Formos Med Assoc. 1997;96(3):217–21.
- Vantrappen G, Feenstra L, Frijns JP. Conductive hearing loss in the trichorhino-phalangeal syndrome (TRP II) or in the Langer-Giedion syndrome. Am J Med Genet. 1997;72(3):372–3.
- Riedl S, Giedion A, Schweitzer K, Mullner-Eidenbock A, Grill F, Frisch H, et al. Pronounced short stature in a girl with tricho-rhino-phalangeal syndrome II (TRPS II, Langer-Giedion syndrome) and growth hormone deficiency. Am J Med Genet A. 2004;131(2):200–3.
- Chen CP, Lin SP, Liu YP, Chern SR, Wu PS, Su JW, et al. An interstitial deletion of 8q23.3-q24.22 associated with Langer-Giedion syndrome, Cornelia de Lange syndrome and epilepsy. Gene. 2013;529(1):176–80.
- Tsang WK, Yang KW, Fong CM. Langer-Giedion syndrome: the evolving imaging features in hands and beyond. Skeletal Radiol. 2014;43(2):251–5.
- Selenti N, Tzetis M, Braoudaki M, Gianikou K, Kitsiou-Tzeli S, Fryssira H. An interstitial deletion at 8q23.1-q24.12 associated with Langer-Giedion syndrome/ Trichorhinophalangeal syndrome (TRPS) type II and Cornelia de Lange syndrome 4. Mol Cytogenet. 2015;8:64.
- Hufeland M, Rahner N, Krauspe R. Trichorhinophalangeal syndrome type I: a novel mutation and Perthes-like changes of the hip in a family with 4 cases over 3 generations. J Pediatr Orthop. 2015;35(1):e1–5.
- Schinzel A, Riegel M, Baumer A, Superti-Furga A, Moreira LM, Santo LD, et al. Long-term follow-up of four patients with Langer-Giedion syndrome: clinical course and complications. Am J Med Genet A. 2013;161A(9):2216–25.
- 14. Ravitch MM, Woods Jr AC. Annular pancreas. Ann Surg. 1950;132(6):1116–27.
- Choi JY, Kim MJ, Kim JH, Lim JS, Oh YT, Chung JJ, et al. Annular pancreas: emphasis on magnetic resonance cholangiopancreatography findings. J Comput Assist Tomogr. 2004;28(4):528–32.
- Hebrok M, Kim SK, St Jacques B, McMahon AP, Melton DA. Regulation of pancreas development by hedgehog signaling. Development. 2000; 127(22):4905–13.
- Jarikji Z, Horb LD, Shariff F, Mandato CA, Cho KW, Horb ME. The tetraspanin Tm4sf3 is localized to the ventral pancreas and regulates fusion of the dorsal and ventral pancreatic buds. Development. 2009;136(11):1791–800.
- Hendricks SK, Sybert VP. Association of annular pancreas and duodenal obstruction-evidence for Mendelian inheritance? Clin Genet. 1991;39(5):383–5.

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