

CASE REPORT

Open Access



Preimplantation genetic testing for Aicardi–Goutières syndrome induced by novel compound heterozygous mutations of *TREX1*: an unaffected live birth

Huiling Xu¹, Jiajie Pu², Suiling Lin¹, Rui Hu¹, Jilong Yao¹ and Xuemei Li^{1*}

Abstract

Background Aicardi–Goutières syndrome (AGS) is a rare, autosomal recessive, hereditary neurodegenerative disorder. It is characterized mainly by early onset progressive encephalopathy, concomitant with an increase in interferon- α levels in the cerebrospinal fluid. Preimplantation genetic testing (PGT) is a procedure that could be used to choose unaffected embryos for transfer after analysis of biopsied cells, which prevents at-risk couples from facing the risk of pregnancy termination.

Methods Trio-based whole exome sequencing, karyotyping and chromosomal microarray analysis were used to determine the pathogenic mutations for the family. To block the inheritance of the disease, multiple annealing and looping-based amplification cycles was used for whole genome amplification of the biopsied trophectoderm cells. Sanger sequencing and next-generation sequencing (NGS)-based single nucleotide polymorphism (SNP) haplotyping were used to detect the state of the gene mutations. Copy number variation (CNV) analysis was also carried out to prevent embryonic chromosomal abnormalities. Prenatal diagnosis was preformed to verify the PGT outcomes.

Results A novel compound heterozygous mutation in *TREX1* gene was found in the proband causing AGS. A total of 3 blastocysts formed after intracytoplasmic sperm injection were biopsied. After genetic analyses, an embryo harbored a heterozygous mutation in *TREX1* and without CNV was transferred. A healthy baby was born at 38th weeks and prenatal diagnosis results confirmed the accuracy of PGT.

Conclusions In this study, we identified two novel pathogenic mutations in *TREX1*, which has not been previously reported. Our study extends the mutation spectrum of *TREX1* gene and contributes to the molecular diagnosis as well as genetic counseling for AGS. Our results demonstrated that combining NGS-based SNP haplotyping for PGT-M with invasive prenatal diagnosis is an effective approach to block the transmission of AGS and could be applied to prevent other monogenic diseases.

Keywords Aicardi–Goutières syndrome, *TREX1*, PGT-M, Monogenic disease

*Correspondence:

Xuemei Li

szfy202204@163.com

Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

Aicardi–Goutières syndrome (AGS) (OMIM 225750) was first described in 1984 by Jean Aicardi and Françoise Goutières [1]. It has a prevalence of 1–5 in 10,000 newly live births [2]. AGS is an autoimmune and genetically heterogeneous disorder with severe neurologic injury [3]. Affected patients' onset usually occurs within the first year of life, and around 25% of them die between the age of 1 and 17 years [4]. AGS primarily affects the brain and the skin. It is most frequently inherited in an autosomal recessive manner and seems to exhibit high variations in penetrance even among patients with same variant [4, 5]. The main clinical features include encephalopathy, leukodystrophy, cerebral atrophy, multiple intracranial calcifications, and lymphocytosis and raised levels of interferon-alpha (IFN α) in the cerebrospinal fluid (CSF) [6]. Nine pathogenic genes associated with AGS have been identified, including *TREX1*, *RNASEH2B*, *RNASEH2C*, *RNASEH2A*, *SAMHD1*, *LSM11*, *RNU7-1*, *ADARI* and *IFIH1* [7, 8].

TREX1 is a 314 amino-acid-long protein encoded by a single exon which is located at 3p21. When preparing this manuscript, there are 197 *TREX1* mutations have been recorded in ClinVar database (<https://www.ncbi.nlm.nih.gov/clinvar/>). *TREX1* is a potent 3'–5' exonuclease that degrades single and double-stranded DNA. When *TREX1* mutates, it will fail to appropriately disassemble genomic DNA during normal cell death processes, which could trigger the innate immune response and autoimmunity [9, 10]. Unfortunately, the main treatment for AGS is only symptomatic. There are still a number of issues regarding AGS treatment needed to be addressed [11, 12].

PGT is a procedure used to analyze gametes or embryos, then unaffected embryos are transferred back to women's uterus, which allow at-risk couples having unaffected children of their own without facing the risk of pregnancy termination [13, 14]. The PGT includes three sub-categories: PGT for aneuploidies (PGT-A), PGT for single monogenic disorders (PGT-M), and PGT for chromosome structural rearrangements (PGT-SR) [15]. In this study, we report a clinically diagnosed AGS caused by a novel compound heterozygous mutation in *TREX1* gene in a Chinese family. These two variants have not been reported previously and have not been recorded as a polymorphic change in public databases [ClinVar (<https://www.ncbi.nlm.nih.gov/clinvar/>), HGMD (<http://www.hgmd.org>), LOVD (<http://www.lovd.nl>)]. To prevent the subsequent transmission of pathogenic mutations in next generation, the couple was counselled and suggested to receive PGT-M treatment. The whole process includes ovarian stimulation, oocyte retrieval, in vitro fertilization, embryo culture and biopsy, and

frozen embryo transfer. Through precise detection procedures, an unaffected embryo was transferred, and a healthy female baby was born at full term.

Methods

Case description

In this case, the Han Chinese none-consanguineous couple were healthy. The mother had her pregnancy at the age of 30. A detailed ultrasound scan at 30 weeks of gestation showed microcephaly and multiple intracranial calcifications. To identify the potential genetic variants through Trio-WES, total genomic DNA was extracted from the whole blood of the couple and amniocytes of the proband. DNA fragments were hybridized and captured by the ClearSeq Inherited Disease Panel (Agilent Technologies, Inc, USA) according to manufacturer's protocol. Then the sequencing library was sequenced on the NovaSeq 6000 platform (Illumina, Inc, USA) with 150 bp paired-end reads. The sequencing reads were aligned to the human reference genome (hg19/GRCh37) with BWA (v0.7.17). Conventional G-banded karyotyping test and CMA test with CytoScan 750 K (Affymetrix, Inc, USA) arrays were performed according to the manufacturer's manual to detect chromosome abnormalities.

In vitro fertilization and blastomere biopsy

The ovarian stimulation, in vitro fertilization, embryo culture and transfer processes were conducted according to the routing protocols [16–18]. In our case, 12 oocytes were obtained and then inseminated by ICSI. Five of them were fertilized normally as indicated by the presence of two pronuclei. In total, 3 blastocysts were obtained. All of them were subjected to TE biopsy on day 5 via laser. Approximately 5–10 cells were extracted from each blastocyst and ready for WGA. The IVF and embryo biopsy were performed at the Center of Reproductive Medicine of the Shenzhen Maternity and Child Healthcare Hospital. WGA and NGS-based PGT were done at Yikon Genomics (Suzhou, China).

PGT with haplotype analysis

Biopsied TE cells from each embryo were applied following the standard protocol of WGA method of MAL-BAC (Yikon Genomics) [19]. The WGA products of each embryo were subjected to Sanger sequencing to identify mutation directly. Because only a few cells are available for amplification, it is hard to avoid allele drop-out (ADO) and is related to the increased risk of DNA contamination. To prevent misdiagnosis, haplotype analysis was conducted through SNP markers. During the pre-testing, SNPs within the 2 Mb region flanking *TREX1* gene are genotyped via NGS using DNA samples from the couple and their parents since the proband's DNA

sample was degraded. Informative SNPs that allow discrimination of the high-risk haplotype and low-risk haplotype are selected for use in the embryo test. A total of thirty SNPs were chosen and screened to determine the possible embryo genotypes via targeted capture sequencing (TCS) with sequencing depth $\geq 100\times$, and specific primers were designed online (<http://www.ampliseq.com>).

CNV analysis

Besides the detection of *TREX1* mutation, CNV analysis was also carried out using WGA products to prevent embryonic abortion, death or other problems may be caused by embryonic chromosomal abnormalities. Any deletions or duplications more than 4Mbp and mosaicism more than 30% in each embryo were reported. Refer to previous study for details [20].

Embryo transfer and prenatal diagnosis

The selection for embryo transfer was based on the morphology and the PGT results. Clinical pregnancy was defined as the presence of a fetal heartbeat by sonography 35 days after frozen embryo transfer (FET). To verify the PGT diagnosis, Sanger sequencing, karyotyping and CMA were conducted using the fetal DNA obtained through amniocentesis at the 17 weeks of gestation.

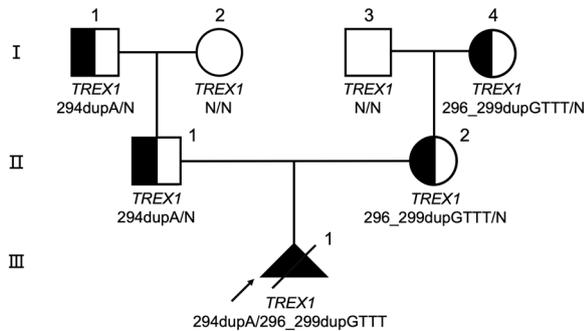


Fig. 1 Pedigree of the AGS family reported in this study. Squares and circles indicate males and females, respectively. The arrowhead denotes the proband

Results

Genetic analysis

As the karyotyping and CMA results showed that the proband had a normal 46, XY karyotype without any deletions or duplications. Trio-WES revealed that the proband harbored a compound heterozygous mutation in *TREX1* gene (maternal: c.296_299dupGTTT, p.Phe100LeufsTer3; paternal: c.294dupA, p.Cys99MetfsTer3) (Fig. 1), which is confirmed by Sanger sequencing. These two mutations were classified as likely pathogenic mutation according to the American College of Medical Genetics and Genomics (ACMG) guideline (PVS1, PM2, PP3) (Additional file 1: Table S1) [21]. Both mutations were predicted to introduce a premature stop codon that affect *TREX1* protein function. Mutation c.294dupA caused the *TREX1* peptide chain shortened from 314 to 202 amino acids. The other mutation c.296_299dupGTTT led to premature translational termination at amino acid position 222, thus the peptide chain was shortened from 314 to 221 amino acids. Pregnancy termination was requested by the parents at the 34 weeks of gestation. Then the couple at risk were underwent PGT treatment.

Embryo preparation and genetic testing

Among the five embryos fertilized by ICSI, three of them (E1, E2, E3) were suitable for biopsy (Table 1). We successfully amplified the DNA of the blastocyst by MALBAC. The CNV results (Fig. 2A, Additional file 3: Fig. S1) showed that only E2 was diagnosed as normal karyotype. Other two embryos showed chromosomal abnormalities related to varying degrees of chromosome mosaicism. After Sanger sequencing (Fig. 2B, Additional file 3: Fig. S1), no mutation was found in *TREX1* for E1, but both E2 and E3 carried the mutation c.294dupA. To verify the Sanger sequencing results, we selected 30 SNPs that could be used for determination of the high-risk haplotype and the low-risk haplotype after pre-testing to infer the disease status of the embryos (Table Additional file 2: S2 and Fig. 3). SNP-based haplotyping showed that both E2 and E3 inherited the paternal high-risk chromosome

Table 1 Detection result summary of the three biopsied blastocysts

Embryo number	Grading#	Sanger sequencing	SNP haplotyping	CNV
E1	3BB	normal	Wild type haplotype	Mixed aneuploidy*
E2	3BB	c.294dupA	Paternal haplotype carrier	46,XN
E3	3CB	c.294dupA	Paternal haplotype carrier	46,XN,4p(pter → p12, ~ 49 Mb, × 1, mos, ~ 50%), 4q(q12 → q35.2, ~ 138 Mb, × 1, mos, ~ 50%)

The embryo quality was assessed following the Gardner grading system [22]

* Embryo with both aneuploid and mosaic chromosomes

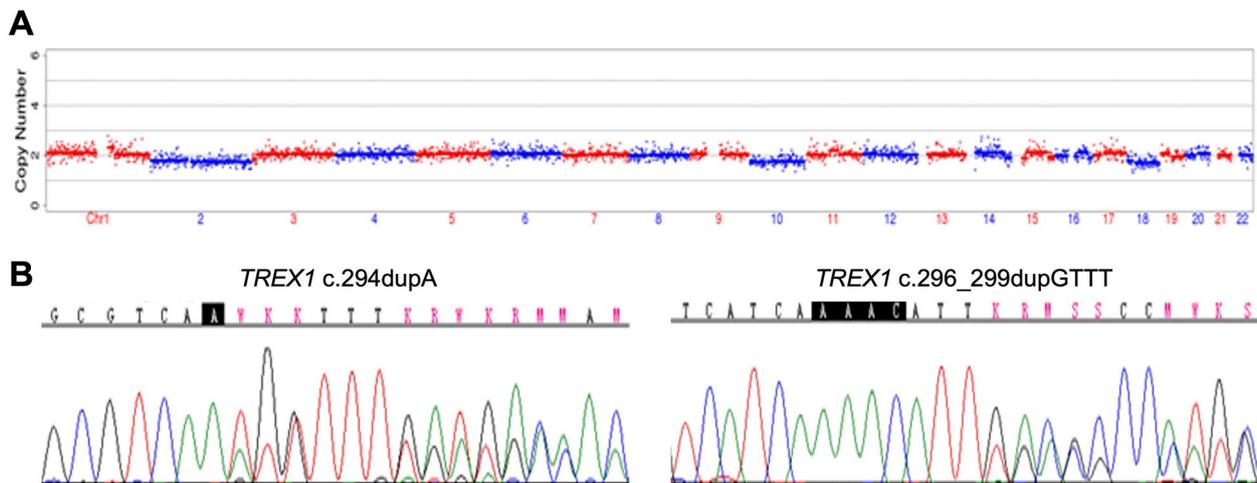


Fig. 2 Copy number variations and Sanger sequencing results of the embryo E2. **A** The sketch map showed that E2 was euploidy. **B** Sanger sequencing result showed that E2 carried the *TREX1* mutation c.294dupA

carrying c.294dupA mutation, while E1 carried neither mutation, which was consistent with the Sanger sequencing.

Prenatal diagnosis and pregnancy outcome

Based on embryo selection principles, the euploid embryo E2 carried heterozygous mutation c.294dupA in *TREX1* was transferred into uterus. During the pregnancy, ultrasound scan showed that the fetus was well developed without microcephaly or intracranial calcifications. The prenatal diagnosis result was consistent with PGT-M. Finally, a healthy female infant was born at the 38th week of gestation.

Discussion

Birth defects bring serious psychological and financial burden to families and have become a crucial social problem. It is estimated that genetic factors contribute solely or collaboratively to about 80% of the occurrence of birth defects. Therefore, genetic studies can provide precise molecular targets for clinical screening, diagnosis and treatment [23]. In this study, we detected two novel missense mutations in *TREX1* gene (c.294dupA and c.296_299dupGTTT) in an AGS fetus, which were inherited from the normal father and mother, separately. Through PGT-M, an embryo was confirmed as euploid as well as heterozygous carrier status of *TREX1* gene, which was verified by Sanger sequencing and haplotype analysis. After transferring the selected embryo, a health fetus was born.

PGT has developed by leaps and bounds since 1989s [24]. It prevents not only the invasive prenatal test which may constitute a risk to the fetus but also the abortion with bearing an affected fetus that may impose psychological and physical burdens on the pregnant woman. However, due to the limited DNA amount of biopsied cells, direct genetic testing through Sanger sequencing based on PCR has the risk of ADO, which is one of the most important causes of the misdiagnoses. SNPs are the most abundant form of human genetic variations, about one SNP every 100–300 bp [25]. And they are easy to interpret and amenable to high-throughput analysis. SNP-based haplotyping could be used to evaluate the occurrence of ADO, monosomy, trisomy and DNA contamination [26]. A clear discrimination between the high-risk and low-risk allele can be made with fully informative SNP markers located close to the target gene. Here, we used NGS-based SNP haplotyping for the verification of the Sanger sequencing results. Such highly informative haplotypes could improve the accuracy and reliability of embryo diagnosis.

Chromosomal mosaicism is at high prevalence in early developmental stage of embryos, which is much higher than reported in the prenatal and postnatal cytogenetic literature [27]. Nowadays, Chromosomal mosaicism is a potential factor of misdiagnosis and has been a raising problem in PGT [28]. A single trophoctoderm biopsy of a few cells couldn't represent the entire embryo. Besides, amplification bias in WGA, sample DNA biological variability, sequencing methods and bioinformatic approaches may lead to false positives. Thirdly, embryo mosaicism may self-correct aneuploidy downstream [29, 30].

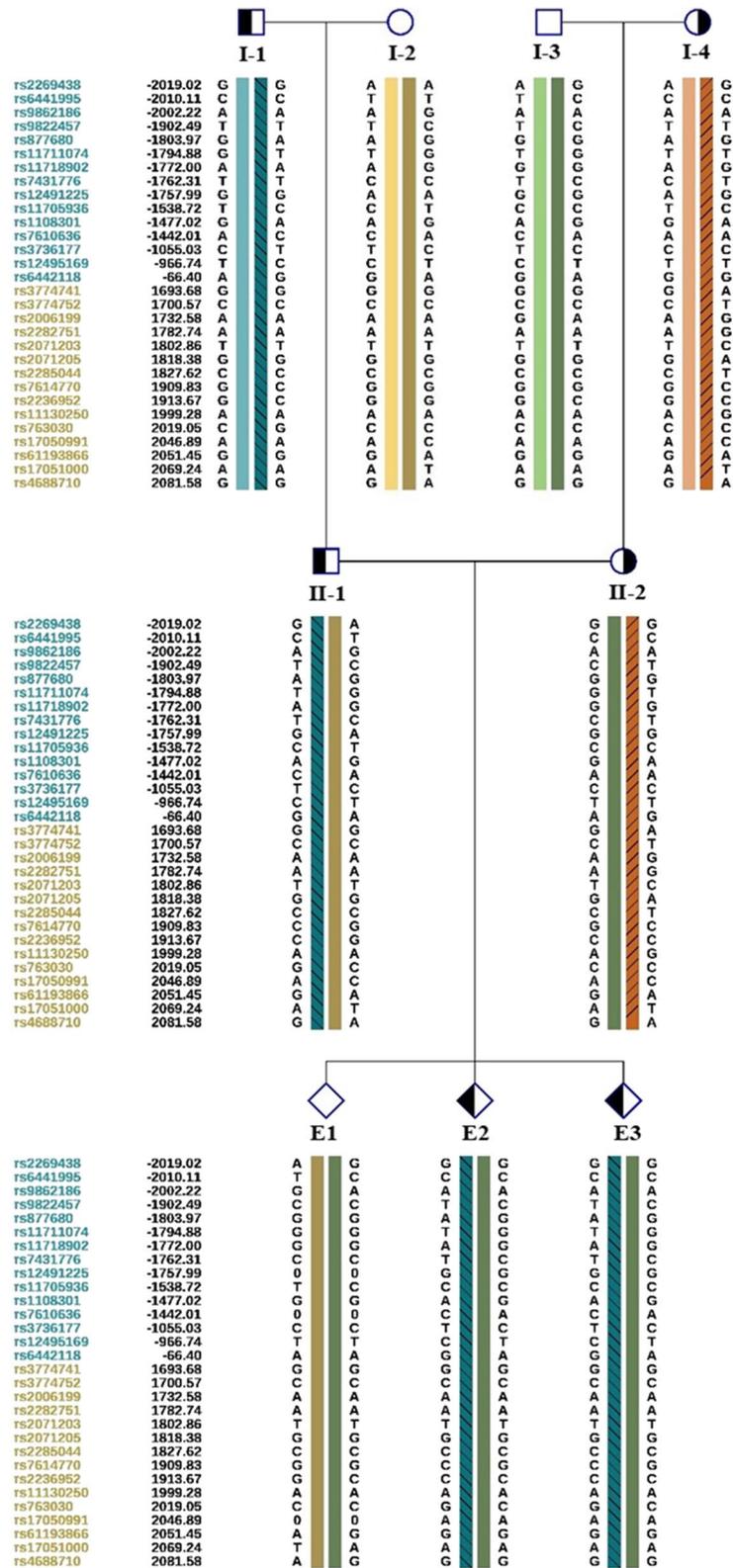


Fig. 3 Pedigree of haplotype linkage analysis. The backslash represents a carrying haplotype inherited from the I-1 and II-1, whereas slash represents a carrying haplotype inherited from I-4 and II-2. Colored rectangle without slash or backslash represents a normal haplotype

Recent studies provided evidence that mosaic embryos can develop into healthy euploid newborns [31–33]. Compared with euploid embryo transfer, however, several studies have reported that mosaic embryo transfer is associated with a lower rate of live birth and a higher rate of biochemical miscarriage [34–36]. Considering the possible risks of transferring mosaic embryos, we eventually selected an euploid carrier embryo E2 to transfer. In this study, rate of mosaicism seems more frequent than reported for that two of the three embryos were mosaic. A possible explanation might be that the use of laser pulse in biopsy improperly increased cell damage which influenced the sequencing results, leading to misdiagnosis of a normal embryo as mosaic. Furthermore, embryo E1 is a complex mosaic blastocyst which involves more than three chromosomes. Recently, a retrospective study showed that 61.6% of the complex mosaic were diagnosed as euploidy in re-biopsy. And the prevalence of the complex mosaic was 2.4% [37]. Therefore, embryos exhibit complex mosaicism, like the E1 in our case, could be re-biopsied to avoid waste of potential available embryos.

We report here our experience of applying NGS-based SNP haplotyping for PGT-M for a couple carrying different mutations in *TREX1* which led to the pregnancy termination of an affected fetus with AGS. According to the age of onset, two clinical presentations of AGS could be delineated: early-onset neonatal form and later-onset presentation. The former highly reminiscent of congenital infection seen particularly with *TREX1* mutations [38]. In our study, 30-week ultrasound scan of the proband showed microcephaly and multiple intracranial calcifications which was initially misdiagnosed as acquired in utero viral infection. *TREX1* playing an important role in processing or clearing anomalous DNA structures, failure of which results in the triggering of an abnormal innate immune response, therefore AGS is sometimes mistaken as the sequelae of congenital infection [10]. Genetic testing is of obvious importance for distinguishing AGS from common perinatal infections.

In conclusion, our study found two novel mutations in *TREX1* gene in a family suffering from AGS. Our study combined NGS-based SNP haplotyping for PGT-M with invasive prenatal diagnosis could effectively block the subsequent transmission of *TREX1* genetic defects, resulting in the birth of a healthy baby. This approach was proven to be practicable while effective and could be expanded to other monogenic diseases.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13039-023-00641-5>.

Additional file 1. Table S1. Variants classification following ACMG guidelines.

Additional file 2. Table S2. NGS-based SNP haplotyping results of the embryo E2.

Additional file 3. Fig S1. Copy number variations and Sanger sequencing results of the embryo E1 and E3.

Acknowledgements

We thank the family for participating in this study.

Author contributions

Devised the idea for this study: LX, YJ; Discussed the results and contributed to the final manuscript: XH, PJ; Obtained patient materials and did clinical operation of PGT: XH, HR; Performed ICSI and the TE biopsy: LS. All authors read and approved the final manuscript.

Funding

The study was carried out with the fund support of the Key Medical Subject of Shenzhen (2020–2024) (SZXK031) and the Research Program of Science, Technology and Innovation Commission of Shenzhen Municipality (2020231557).

Availability of data and materials

The datasets generated and/or analyzed during the current study are not publicly available due to individual privacy or ethical restrictions but are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of Shenzhen Maternity and Child Healthcare Hospital of Southern Medical University (No. 2021-86). All patients provided written informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Reproductive Medicine, Southern Medical University Affiliated Shenzhen Maternity and Child Healthcare Hospital, Shenzhen, Guangdong, China. ²Department of Bioinformatics, 01life Institute, Shenzhen 518000, Guangdong, China.

Received: 1 February 2023 Accepted: 25 May 2023

Published online: 05 June 2023

References

1. Aicardi J, Goutières F. A Progressive familial encephalopathy in infancy with calcifications of the basal ganglia and chronic cerebrospinal fluid lymphocytosis: aicardi and Goutières: Basal Ganglia Calcifications. *Ann Neurol*. 1984;15:49–54.
2. Viguera Elías D, De La Iglesia NI, Toledo Gotor C, Domínguez Garrido E, Poch Olivé ML. Síndrome de Aicardi–Goutières: un caso familiar por alteración del gen RNASEH2B. *RevNeurol*. 2021;72:407.

3. Sase S, Takanohashi A, Vanderver A, Almad A. Astrocytes, an active player in Aicardi–Goutières syndrome: astrocytes in Aicardi–Goutières syndrome. *Brain Pathol.* 2018;28:399–407.
4. Chahwan C, Chahwan R. Aicardi–Goutières syndrome: from patients to genes and beyond. *Clin Genet.* 2012;81:413–20.
5. Zhang S, Song J, Yang Y, Miao H, Yang L, Liu Y, et al. Type I interferonopathies with novel compound heterozygous TREX1 mutations in two siblings with different symptoms responded to tofacitinib. *Pediatr Rheumatol.* 2021;19:1.
6. Hutchinson AK, Ngo H, Silverstein E, Couser NL. Genetic abnormalities of ocular motility. In: *Ophthalmic genetic diseases.* Elsevier; 2019. p. 107–16.
7. Crow YJ, Chase DS, Lowenstein Schmidt J, Szykiewicz M, Forte GMA, Gornall HL, et al. Characterization of human disease phenotypes associated with mutations in TREX1, RNASEH2A, RNASEH2B, RNASEH2C, SAMHD1, ADAR, and IFIH1. *Am J Med Genet.* 2015;167:296–312.
8. Uggenti C, Lepelley A, Depp M, Badrock AP, Rodero MP, El-Daher M-T, et al. cGAS-mediated induction of type I interferon due to inborn errors of histone pre-mRNA processing. *Nat Genet.* 2020;52:1364–72.
9. Fye JM, Orebaugh CD, Coffin SR, Hollis T, Perrino FW. Dominant mutations of the TREX1 exonuclease gene in lupus and Aicardi–Goutières syndrome. *J Biol Chem.* 2011;286:32373–82.
10. Crow YJ, Hayward BE, Parmar R, Robins P, Leitch A, Ali M, et al. Mutations in the gene encoding the 3′-5′ DNA exonuclease TREX1 cause Aicardi–Goutières syndrome at the AGS1 locus. *Nat Genet.* 2006;38:917–20.
11. Tonduti D, Fazzi E, Badolato R, Orcesi S. Novel and emerging treatments for Aicardi–Goutières syndrome. *Expert Rev Clin Immunol.* 2020;16:189–98.
12. Crow YJ, Shetty J, Livingston JH. Treatments in Aicardi–Goutières syndrome. *Dev Med Child Neurol.* 2020;62:42–7.
13. Kuliev A, Rechitsky S. Preimplantation genetic testing: current challenges and future prospects. *Expert Rev Mol Diagn.* 2017;17:1071–88.
14. Simpson JL, Kuliev A, Rechitsky S. Overview of preimplantation genetic diagnosis (PGD): Historical perspective and future direction. In: Levy B, editor. *Prenatal Diagnosis.* New York: Springer; 2019. p. 23–43.
15. Fesahat F, Montazeri F, Hoseini SM. Preimplantation genetic testing in assisted reproduction technology. *J Gynecol Obstet Hum Reprod.* 2020;49:101723.
16. McArthur SJ, Leigh D, Marshall JT, De Boer KA, Jansen RPS. Pregnancies and live births after trophoctoderm biopsy and preimplantation genetic testing of human blastocysts. *Fertil Steril.* 2005;84:1628–36.
17. The ESHRE Guideline Group on Ovarian Stimulation, Bosch E, Broer S, Griesinger G, Grynberg M, Humaidan P, et al. ESHRE guideline: ovarian stimulation for IVF/ICSI†. *Hum Reprod Open.* 2020;2020:hoaa009.
18. ESHRE PGT Consortium and SIG-Embryology Biopsy Working Group, Kokkali G, Coticchio G, Bronet F, Celebi C, Cimadomo D, et al. ESHRE PGT Consortium and SIG Embryology good practice recommendations for polar body and embryo biopsy for PGT†. *Hum Reprod Open.* 2020;2020:hoaa020.
19. Zong C, Lu S, Chapman AR, Xie XS. Genome-wide detection of single-nucleotide and copy-number variations of a single human cell. *Science.* 2012;338:1622–6.
20. Bai H, Li X, Liu X, Shi W, He B, Wei R, et al. Preimplantation genetic testing for recurrent autosomal dominant osteogenesis imperfecta associated with paternal gonosomal mosaicism. *Front Genet.* 2022;13:1011833.
21. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med.* 2015;17:405–24.
22. Balaban B, Gardner DK. Morphological assessment of blastocyst stage embryos: Types of grading systems and their reported outcomes. In: Gardner DK, Sakkas D, Seli E, Wells D, editors. *Human gametes and preimplantation embryos.* New York: Springer; 2013. p. 31–43.
23. Sun LY, Xing QH, He L. Retrospect and prospect of the genetic research on birth defects in China [in Chinese]. *Yi Chuan = Hereditas.* 2018;40:800–13.
24. Handyside AH, Kontogianni EH, Hardy K, Winston RML. Pregnancies from biopsied human preimplantation embryos sexed by Y-specific DNA amplification. *Nature.* 1990;344:768–70.
25. Marth GT, Korf I, Yandell MD, Yeh RT, Gu Z, Zakeri H, et al. A general approach to single-nucleotide polymorphism discovery. *Nat Genet.* 1999;23:452–6.
26. ESHRE PGT-M Working Group, Carvalho F, Moutou C, Dimitriadou E, Dreesen J, Giménez C, et al. ESHRE PGT Consortium good practice recommendations for the detection of monogenic disorders†. *Human Reproduction Open.* 2020;2020:hoaa018.
27. Zhang YX, Chen JJ, Nabu S, Yeung QSY, Li Y, Tan JH, et al. The pregnancy outcome of mosaic embryo transfer: a prospective multicenter study and meta-analysis. *Genes.* 2020;11:973.
28. Takeuchi K. Pre-implantation genetic testing: Past, present, future. *Reprod Med Biol.* 2021;20:27–40.
29. Orvieto R. The reproducibility of trophoctoderm biopsies – The chaos behind preimplantation genetic testing for aneuploidy. *Eur J Obstet Gynecol Reprod Biol.* 2020;254:57–8.
30. Gleicher N, Patrizio P, Brivanlou A. Preimplantation genetic testing for aneuploidy—a castle built on sand. *Trends Mol Med.* 2021;27:731–42.
31. Greco E, Minasi MG, Fiorentino F. Healthy babies after intrauterine transfer of mosaic aneuploid blastocysts. *N Engl J Med.* 2015;373:2089–90.
32. Munné S, Spinella F, Grifo J, Zhang J, Beltran MP, Fragouli E, et al. Clinical outcomes after the transfer of blastocysts characterized as mosaic by high resolution Next Generation Sequencing—further insights. *Eur J Med Genet.* 2020;63:103741.
33. Capalbo A, Poli M, Rienzi L, Girardi L, Patassini C, Fabiani M, et al. Mosaic human preimplantation embryos and their developmental potential in a prospective, non-selection clinical trial. *Am J Hum Genet.* 2021;108:2238–47.
34. Fragouli E, Alfarawati S, Spath K, Babariya D, Tarozzi N, Borini A, et al. Analysis of implantation and ongoing pregnancy rates following the transfer of mosaic diploid–aneuploid blastocysts. *Hum Genet.* 2017;136:805–19.
35. Zhang L, Wei D, Zhu Y, Gao Y, Yan J, Chen Z-J. Rates of live birth after mosaic embryo transfer compared with euploid embryo transfer. *J Assist Reprod Genet.* 2019;36:165–72.
36. Zore T, Kroener LL, Wang C, Liu L, Buyalos R, Hubert G, et al. Transfer of embryos with segmental mosaicism is associated with a significant reduction in live-birth rate. *Fertil Steril.* 2019;111:69–76.
37. Zhou S, Xie P, Zhang S, Hu L, Luo K, Gong F, et al. Complex mosaic blastocysts after preimplantation genetic testing: prevalence and outcomes after re-biopsy and re-vitrification. *Reprod Biomed Online.* 2021;43:215–22.
38. Rice G, Patrick T, Parmar R, Taylor CF, Aeby A, Aicardi J, et al. Clinical and molecular phenotype of Aicardi–Goutières syndrome. *Am J Hum Genet.* 2007;81:713–25.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

